RESEARCH ARTICLE

Exploring the Hidden Potential of Traditional Rice Landraces and Mutants for Iron and Zinc Concentration in Brown Rice to Develop Bio-fortified Rice Varieties

Deepika Parte¹, Parmeshwar K. Sahu¹, Ravi Raj S. Patel¹, Richa Sao¹, Samrath Baghel¹, Kuber Bhainsa², B.K. Das² and **Deepak Sharma1***

Abstract

In the present study, 58 rice genotypes were evaluated for three consecutive seasons to enucleate the elite lines with high Fe and Zn contents. ANOVA revealed significant differences among the genotypes for Fe and Zn contents during all three seasons. Furthermore, PCV and GCV were moderate to high for Fe and Zn content during all three seasons. Similarly, high level of heritability and genetic advance were also recorded for both the traits. Iron content of Safri mutant 17-2-48 was consistently high for all the seasons whereas zinc content of Loktimachi was high for two seasons. Interestingly, rice genotypes Chhindmauri, Badsahbhog, Rudra, Danighoda and Suapankhi have been identified to have moderate to high Fe and Zn contents which could be used as donors for developing bio-fortified rice varieties. Stability analysis revealed that the genotypes Jhimiprass, Loktimachi, and CG Zinc Rice 2 for iron content and Rela Dhan, Chhindmauri, and CG Zinc Rice-1 for zinc content have had an average response to environment, making them appropriate for all seasons **Keywords**: Rice landraces, Bio-fortification, Fe & Zn content, Variability, Stability analysis.

1 Department of Genetics and Plant Breeding, Indira Gandhi Krishi Vishwavidyalaya (IGKV), Raipur, Chhattisgarh, India.

2 Nuclear Agriculture and Biotechnology Division, Bhabha Atomic Research Centre (BARC), Mumbai, Maharashtra, India.

***Author for correspondence:**

deepak1962@igkv.ac.in

Received:20/09/2022 **Revised:**20/02/2023

Accepted:22/05/2023

How to cite this article: Parte, D., Sahu, P.K., Patel, R.R.S., Sao, R., Baghel, S., Bhainsa, K., Das, B.K., Sharma, D. (2023). Exploring the Hidden Potential of Traditional Rice Landraces and Mutants for Iron and Zinc Concentration in Brown Rice to Develop Bio-fortified Rice Varieties. *Indian J. Plant Genetic Resources*. 36(2), 280-289. **DOI:**10.61949/0976-1926.2023.v36i02.10

Introduction

Rice is a major staple food for more than half of the world's population and contributes to solve the problem of hunger among poor people, but they are rarely accessible to nutrient rich food sources. The prevalence of micronutrient deficiency, especially for crucial elements like Fe content and Zn content among the rice-consuming population, prompted in initiation bio-fortification programme in rice which has been identified as a sustainable source to develop nutrient-rich rice grains to poor people who depend solely on rice for the energy and nutrients (Rao *et al*., 2014). Traditional rice landraces have tremendous genetic variability and possess valuable genes for nutritional traits. Several biotic and abiotic stresses have adaptability in wide agroecological niches, unmatched qualitative traits and therapeutic properties, and thus possesses great genetic potential for rice improvement. Along with the rice landraces, mutants developed from them are a valuable source of micronutrients that need to be quantified and utilized to develop bio-fortified rice varieties. Production of varieties containing high amounts of bioavailable Fe would improve Fe nutrition in regions with prevalent iron deficiency (Aggett, 2020; Maret and Sandstead, 2006; Shahzad *et al*., 2014). Studies by Harvest Plus and others have shown considerable iron and zinc

© IJPGR, 2023. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit https://creativecommons.org/licenses/by-nc-sa/4.0/. losses during rice polishing. Therefore, an increase in the concentration of Fe and Zn in brown rice is a high-priority research area and later on that could be quantified in white rice for varietal development (Ryu and Aydemir, 2020). For this concern, we need to focus on micronutrient-rich sources which can be easily reachable and acceptable to people.

Biofortification is one of the sustainable approaches for improving the iron and zinc content and their bioavailability in rice grains. Nevertheless, before the implementation of the biofortification process, the primary step in conventional breeding is to screen out the micronutrient-dense cultivars within the natural existing germplasm because the presence of awide range of genetic variations in the population helps in selecting the desired genotypes. Furthermore, selecting stable genotypes for high micronutrient content is also crucial for developing nutri-rich rice varieties. Therefore, the current study utilized 44 rice landraces and 8 mutants along with the 6 check to explore the genetic variability, stability and hidden potential of traditional rice landraces and mutants for iron and zinc concentration in brown rice over different seasons. Moreover, the outcome of this study will be further helpful to create the genetic database for breeding program specific for improving the micronutrient in the rice varieties.

Materials and Methods

Experimental Site

The field experiments were conducted at Instructional Farm, Indira Gandhi Krishi Vishwavidyalaya, Raipur-492012 (C.G.) India, during three seasons *viz.*, *Kharif* season*-*2020, *Rabi* season *-*2020-21, and *Kharif* season *-*2021. Geographically, Chhattisgarh state of India is located between 17°14' N and 24°45' N latitude and longitude of 79°16' E. The experiment was laid out by following a randomized complete block design with two replications during all three seasons. All the standard agronomic practices were adopted during the crop seasons.

Plant Materials

A total of 58 accessions, including 44 landraces, 08 mutants, and 06 checks, were used in the experiment. All the genotypes were accessed from the Department of Genetics and Plant Breeding, Indira Gandhi Krishi Vishwavidyalaya, Raipur-492012 (C.G.). The list of experimental materials is presented in Table 1. Dehusked brown rice was used to estimate Zinc and Iron content during all three seasons.

Estimation of Iron and Zinc Contents

Iron and zinc content were estimated using the S2 RANGER ED-XRF Method at International Rice Research Institute, South Asia Hub, ICRISAT Campus, Hyderabad, India center. The seed samples were dehusked by the help of Zaccaria testing rice mill having rubber roller. Before dehusking,

the grains were washed with 0.1N HCl followed by rinsing with double distilled water to make the sample free from contamination. Total 5 g of properly cleaned brown rice in triplicates were used for the estimation of Fe and Zn contents. The published protocol of ED-XRF was used to estimate both the micro-nutrients (Rao *et al.,* 2014). The data from the application was exported in MS Excel sheet for further analysis.

Table 1: List of rice genotypes used as experimental materials under the study

u ic stuuy							
S. No.	Name of genotypes	Parentage					
1	Red Barhasal	landrace					
2	Rudra	landrace					
3	Tulsibhog	landrace					
4	Loktimachi	landrace					
5	Odha Dhan	landrace					
6	Gathuwan Dhan	landrace					
7	Raja Bangla	landrace					
8	Brown Rice-1	landrace					
9	Munibhog	landrace					
10	Danighoda	landrace					
11	Saraitolia	landrace					
12	Pangudi Goindi	landrace					
13	Safri-17	landrace					
14	Dubraj	landrace					
15	Byalo	landrace					
16	Badshahbhog	landrace					
17	Jonyaphool	landrace					
18	Mohlyanbanko	landrace					
19	Kanakbhog	landrace					
20	Shwet Ganga	landrace					
21	Matko Dhan	landrace					
22	Suapankhi	landrace					
23	Jhimiprass	landrace					
24	Jauphool	landrace					
25	Kalajeera	landrace					
26	Chhindmauri	landrace					
27	Motipeera	landrace					
28	Pedgadi	landrace					
29	Sonagathi	landrace					
30	Jana Dhan	landrace					
31	Rela Dhan	landrace					
32	Ram Shree	landrace					
33	Kari Alcha	landrace					
34	Jhilli Safri	landrace					
35	Siyar	landrace					
36	Karhani	landrace					

Statistical Analysis

Statistical analysis for analysis of variance (ANOVA), genetic variability parameters like mean, range, phenotypic coefficients of variation (PCV), genotypic coefficients of variation (GCV), heritability in broad sense (Hb), and genetic advance as percent of the mean parameters were calculated through the standard software (Windostat version 9.2 from Indostat services, by following the standard methodology. Stability analysis over the three seasons was performed following the Eberhart and Russell (1966) Model through Windostat version 9.2 from Indostat services.

Results

Total 58 accessions were analyzed and the result of pooled variance analysis (Table 2) revealed high variability for the iron and zinc as represented by the highly significant mean sum of squares over three seasons. However, the results of **Table 2:** Pooled analysis of variance (ANOVA) for Fe and Zn contents over the three seasons (*Kharif season* 2020, Rabi season 2020-21, and *Kharif season* 2021.)

** Significant at 1% level of significance, * Significant at 5% level of significance

different genetic variability parameters revealed that the phenotypic coefficients of variation (PCV) were slightly higher than the corresponding genotypic coefficients of variation (GCV) for both traits among all three seasons (Table 3). Heritability in broad sense (Hb) were found to be medium for iron (36.30%) and zinc (41.78%) in season I while in season II it was estimated high for Iron (83.54%) and Zinc (80.21%). In season III heritability was high for Zn (73.17%) but found medium for Fe (64.68%). Similarly, high estimate of genetic advance as a percent of mean was recorded for Fe in season II (39.92) and season III (27.3), while it was estimated as medium in season I (16.66). However, genetic advance as a percent of mean for Zn was estimated to be medium in all three seasons (Table 3). Total 58 accessions were analyzed and variation in iron and zinc contents in brown rice was studied in different seasons.

In season I, range for the concentration of iron varied from 8.13 \pm 0.46 to 20.83 \pm 0.71 ppm with an average of 11.41 ppm iron, while zinc content among the 58 genotypes ranged from 14.41 \pm 1.29 to 27.45 \pm 1.45 ppm with an average of 21.41 ppm zinc. Among all genotypes, Safri mutant 17-2-48 had the highest iron content (20.83 ppm), while Red Barhasal recorded the lowest iron content during *Kharif*-2020.

Meanwhile, Munibhog recorded the highest zinc content and Odha Dhan recorded the least (Table 4). Among the genotype tested, 78% were found high for iron content while 22% of genotypes had low iron content, 74% contained high zinc, and 26% had low zinc content (Figure 1) in season I. The top five elite genotypes with high iron during season I are Safri-17-2-48 Mutant, Badshahbhog, Rajeshwari, Chhindmauri, and Rudra (Table 5). The top five elite

Table 3: Genetic variability parameters for Fe and Zn contents over the three seasons

Micronut rients	Kharif season 2020			Rabi season 2020-21			Kharif season 2021								
	lear	88	န္က ဥ		$\begin{array}{ccc}\n\text{Mean} & \text{G} & \text{M} \\ \text{G} & \text{M} & \text{M} \\ \text{M} & \text{M} & \text{M}\n\end{array}$		* ಗ ″	* ၅		$C \underset{\text{gen}}{\approx} C$	Mea	\sum_{∞}	* ၅	ㅎ	GA % mear
Fe	11.42	22.29	13.43	36.30	16.66	11.85 23.2		21.21	83.54 39.92		11.63	20.47	16.46	64.68	27.3
Zn.	21.42	15.69	10.14	41.78	13.51	20.93	12.47	11.17	80.21	20.61	21.18	12.58	10.76	73.17	- 19

genotypes with high zinc are Munibhog, Byalo, Jhimiprass, Jonyaphool, and Loktimachi (Table 6). Iron concentration in season II ranged from 8.31 ± 0.01 ppm to 22.19 ± 2.63 ppm and zinc concentration from 16.33 ± 1.03 to 25.80 ± 0.01 ppm with the mean value of 11.85 and 20.93 ppm for iron and zinc respectively (Table 4). Meanwhile, out of tested genotypes, 83% of total genotypes had high iron content while 17% of genotypes have low iron content, whereas for a concentration of zinc 67% of genotypes contained high zinc while 33% has low zinc in brown rice (Figure 2).

The top elite genotypes with high iron during season II are Safri-17-2-48 Mutant, TCDM-1, Rudra, Chhindmauri, and Suapankhi, respectively (Table 5). The top elite genotypes with high zinc are Loktimachi, TCDM-1, Jhimiprass, Badshahbhog, and Chhindmauri (Table 6). Chhindmauri and TCDM-1 can be used as the donor parent for both iron and zinc.

However, variation in iron during the third season ranged from 8.68 ± 0.35 (Raja Bangla) to 21.51 ± 0.96 ppm (Safri-17-2-48 Mutant) and zinc concentration from 15.38 \pm 1.37 (Odha Dhan) to 25.80 \pm 0.01 ppm (Loktimachi) with the mean value of 11.63 ppm for iron and 21.18 ppm for zinc (Table 4). Out of 58 genotypes, 84% found high iron while 16% have low iron content, 67% contain high zinc while 33% have low zinc, as shown in Figure 3. The top five genotypes with high iron during season III are Safri 17-2-48 Mutant, Rudra, Chhindmauri, TCDM-1, and Badshahbhog (Table 5). While the top five genotypes which having high zinc are Loktimachi, Byalo, Munibhog, Jhimiprass, and Chhindmauri (Table 6) these genotypes can be utilized to develop biofortified varieties.

Among all the seasons, the concentration of Iron was found high for Safri mutant 17–2-48, where Munibhog recorded for highest zinc concentration during season I, whereas Loktimachi found the highest during the second and third seasons. Thus, variations in rice grains' iron and zinc content were observed during different seasons in the present study. These variations can be attributed to different factors such as environment, soil properties, nature of grain, genotype, and interaction of genotype X environment, micronutrient homeostasis, method of analysis, *etc*. Furthermore, Chhindmauri, Badshahbhog, Rudra, Danighoda, Suapankhi, TCDM-1 and Luchai Mutant-1 exhibited best for both Zn and Iron. Thus, these genotypes can be used as donor parents for developing Iron and Zinc rich varieties.

To test the significance among the genotypes, mean sum of square due to genotypes has been tested against mean sum of square due to pooled deviation. The results indicated highly significant mean sum of square due to genotypes for both iron and zinc contents in brown rice, indicating the availability of ample variability among the genotypes (Table 7). Stability analysis revealed that all the genotypes included in the study deviated non-significantly from zero

Figure 1: Percentage of genotypes having high and low Iron and Zinc content during *Kharif* season 2020 (season I)

Figure 2: Percentage of genotypes having high and low Iron and Zinc content during *Rabi* season 2020-21 (season II)

Figure 3: Percentage of genotypes having high and low Iron and Zinc content during *Kharif* season 2021 (season III)

 $(\sigma^2 d=0)$. Still, only genotypes Jhimiprass, Loktimachi and CG zinc rice two deviated non-significantly from unity of regression for iron content. The 3 genotypes *viz*., Rela Dhan, Chhindmauri and CG Zinc Rice 1 were deviated non-significantly from unity of regression for zinc content (Table 8). Hence these genotypes have an average response to environments and are recommended for all seasons (environments), while the regression coefficient of 32 genotypes for iron and 27 genotypes for zinc recorded significant deviation from unity and less than unity (b<1). Hence these genotypes have below average response to environments and are recommended for poor unfavorable environments. Furthermore, the regression coefficient of 23 genotypes for iron and 25 genotypes for zinc recorded significant deviation from unity and more than unity (b > 1). Therefore, it has an above-average response to environments and is recommended for favorable environments.

Discussion

Iron and zinc deficiency is notably the most widespread micronutrient deficiency among the human population. As a staple food, a lot of efforts are being made to enrich the nutritional status of rice to prevent malnutrition. However, the effect of these efforts is still limited because of the low consumer acceptability of fortified food and the low bioavailability of iron and zinc in our resources (Pyo *et al.,* 2022). For this concern, knowledge of our germplasm resources and their preferentially elemental constitutions of the different grain tissues and their remobilization patterns during germination makes the possibility of designing target products with a nutritionally optimal constitution more feasible (Arora *et al*., 2010; Omary *et al*., 2012). While a better understanding of the physiological basis of nutrient

Table 4: Mean performance of rice landraces and mutant for Fe and Zn contents over the three seasons

Deepika Parte *et al.* **Genetic variation and adaptation study for micro-nutrients in rice landraces and mutants**

Table 5: Top performing genotypes for Fe contents in rice genotypes

Table 6: Top performing genotypes for Zn contents in rice genotypes

uptake (Kong *et al*., 2022), their translocation, and the maintenance of homeostasis in grain is essential for the genetic biofortification of rice, complete knowledge of these processes in rice is still lacking (Olsen and Palmgren, 2014; Kobayashi *et al.,* 2021).

Ultimately, the concentration of micronutrients in rice grain depends on the genotype's efficiency to uptake and translocate the nutrients from root to the grain, which involves many complex physiological processes at different levels. Therefore, rice genotypes to be developed should have the genetic potential and physiological efficiency to utilize the available micronutrients from the soil (Olsen and Palmgren, 2014). Knowledge of mineral localization

within rice grains is important for understanding the role of different elements in seed development and facilitating the biofortification of seed micronutrients to enhance seeds' values in human diets (Lu *et al.*, 2013).

To initiate a breeding program to improve or develop any variety depends on the availability of variation in germplasm for the targeted trait, which can be further used in genetic studies, to make crosses, molecular marker development, and to understand the basis of the uptake process for micronutrients. A large genetic variation for grain iron and zinc has been observed in different germplasm of rice and it was exploited in breeding programs. In our study Iron content in season I, ranged between 8.13 \pm 0.46 to

 20.83 ± 0.71 ppm and zinc content between 14.41 \pm 1.29 to 27.45 ± 1.45 ppm in brown rice. Similarly, in season II, Iron content ranged from 8.31 ± 0.01 to 22.19 \pm 2.63 ppm and zinc from 16.33 \pm 1.03 to 25.80 \pm 0.01 ppm. Interestingly, the range of micronutrients were slightly higher in season III, *viz*., 8.68 ± 0.35 to 21.51 ± 0.96 ppm for iron content and 15.38 ± 1.37 to 25.80 ± 0.01 ppm for zinc content. Zhang *et al.,* (2018) experimented on a set of 698 rice germplasms and found that the iron concentration ranged between 0.9 to 9.1 μg/g while zinc concentration ranged from 5.8 to 29.6 μg/g in the polished rice. Similarly, Roy and Sharma (2014) found ample variation for Fe and Zn content in 84 rice cultivars and reported that Iron content varied between 0.25 to 34.8 μg/g and zinc content from 0.85 to 195.3 μg/g

> where local cultivars had the highest iron content. The grain iron content was found to be relatively higher in the second season with the maximum values of 22.19 ± 2.63

Table 7: Analysis of variance for stability of iron and zinc content among rice genotypes

in the current study. However, the seasonal variations for these elements were observed as reported earlier (Dixit *et al.,* 2019; Suman *et al.,* 2021). The variation in values may arise due to properties of grain such us relative position on the panicle (Su *et al.*, 2014), moisture content (Rao *et al*., 2014), number of aleurone layer (Sellappan *et al*., 2009), etc.; soil properties such as pH, organic carbon content and inherent availability of iron and zinc in the soil (Chandel *et al*., 2010). Furthermore, high significant interaction effect of genotype x environment on the content of these micronutrients has been reported by Chandel *et al.* (2010); and Suwarto (2011). In the current study, iron and zinc contents were found high in landraces and similar results were observed and reported earlier (Anandan *et al.,* 2011; Nachimuthu *et al.,* 2014; Parikh *et al.*, 2019). Sanjeeva *et al.,* (2020) experimented on various sets of germplasm and reported that 99 genotypes from

germplasm and 344 lines from mapping populations exhibited≥ 28 mg/kg zinc content in polished rice, meeting the target zinc content set by HarvestPlus.

In this study, Safri mutant 17-2-48 has the highest iron content, a newly developed mutant from Safri 17 local variety. This mutant can be directly released as a new variety with high iron content. Munibhog and Loktimachi were identified for the highest zinc content. Thus, these genotypes open the possibilities for the exploitation as a donor in biofortification breeding programs and the identification of genomic positions associated with iron and zinc contents in grains. Though the variation in micronutrient content depends on several factors, the germplasm stock in rice has sufficient variation to exploit for developing stable lines. As micronutrient malnutrition poses a significant global challenge, the development of micronutrient enriched genotypes serves as the need of the hour. The top-performing genotypes identified in this research will be useful for selecting and breeding lines with enriched micronutrient status.

A stable variety for grain nutrition quality is an important consideration for biofortification. The genotype, environment, linear genotype X environment and nonlinear genotype X environment interaction when tested against pooled deviation, showed significance for both iron and zinc content in the brown rice. Similar significance for zinc and iron content have earlier been reported by Oikeh *et al.*, (2004), Prasanna *et al*., (2011), Suwarto (2011), Velu *et al*., (2012) and Ajmera *et al*., (2017). For iron content the genotypes Jhimiprass, Loktimachi and CG Zinc Rice 2 did not deviate from unity of regression, showing their stable and predictable performance. Therefore, they can be recommended for growing in all environments ranging from optimum to marginal environments. Similarly, for Zinc content the genotypes Rela Dhan, Chhindmauri and CG Zinc Rice 1 did not deviate significantly from the unity of regression. Therefore, these genotypes can be grown in any environment to get more or less stable performance in terms of the Zinc content in brown rice. Genotypes 32 and 27 recorded significant deviation from unity and less than unity (b< 1) for Iron and Zinc content, respectively, showing their below-average performance. These genotypes cannot take full advantage of the favorable conditions when grown in the optimum environment. Therefore, they are suitable for cultivation in marginal or suboptimal environments such as *Rabi* season. However, 23 and 25 genotypes for iron and zinc content have recorded significant deviation from unity and more than unity (b > 1). When grown in the optimum environment with the most suitable condition, such genotypes will result in above-average performance in terms of the micronutrient content in brown rice. Therefore, to take the best advantage, these varieties can be grown in *Kharif* season at regions having good soil fertility.

Conclusion

Existing genetic variation in rice germplasm offers scope for developing nutrient-rich varieties. Notably, there was an ample difference in Fe and Zn content suggesting the existence of genetic potential to increase the concentration of these micronutrients in rice grain. Micronutrient-rich rice genotype *viz*., Safri-17-2-48 Mutant, Loktimachi, Chhindmauri, Badsahbhog, Rudra, Danighoda, Suapankhi, TCDM-1, and Luchai Mutant-1 identified through this study may be further utilized in the breeding program as a donor for development of nutrient-rich or biofortified rice varieties. Additionally, genotypes Jhimiprass, Loktimachi, and CG zinc Rice 2 for iron content and Rela Dhan, Chhindmauri, and CG zinc rice 1 for zinc content exhibited an average response to the environment, making them suitable for all seasons.

Acknowledgment

We sincerely thank IRRI-SA Hub, ICRISAT campus, Hyderabad, India for providing the facility for iron and zinc estimation.

References

- Aggett PJ (2020) Iron. In: Present Knowledge in Nutrition (Eleventh Edition), BP Marriott, DF Birt, VA Stallings and AA Yates (Ed.), Academic Press, USA, p 375–392.
- Ajmera S, SS Kumar and V Ravindrababu (2017) Genotype × Environment interactions and stability analysisfor grain iron and zinc concentrations in Rice (*Oryza Sativa* L.)Genotypes. *Int. J. Curr. Microbiol. App. Sci.* **6(7):** 1902-1913.
- Anandan A, G Rajiv, R Eswaran, and M Prakash (2011) Genotypic variation and relationships between quality traits and trace elements in traditional and improved rice (*Oryza sativa* L.) genotypes. *J. Food Sci*. **76(4):** H122–H30.
- Arora S, S Jood, and N Khetarpaul (2010) Effect of germination and probiotic fermentation on nutrient composition of barley-based food mixtures. *Food Chem.* **119(2):** 779-784.
- Chandel G, S Banerjee, S See, R Meena, DJ Sharma and SB Verulkar (2010) Effects of different nitrogen fertilizer levels and native soil properties on rice grain Fe, Zn and protein contents. *Rice Sci*. **17(3):** 213−227.
- Dixit S, UM Singh, R Abbai, T Ram, VK Singh and A Paul (2019) Identification of genomic region(s) responsible for high iron and zinc content in rice. *Sci. Rep*. **9:** 8136. doi: 10.1038/ s41598-019-43888-y
- Eberhart SA and WA Russell (1966) Stability parameters for comparing varieties. *Crop Sci*. **6(1):** 36- 40.
- Kobayashi T, AJ Nagano and NK Nishizawa (2021) Iron deficiency‐ inducible peptide‐coding genes OsIMA1 and OsIMA2 positively regulate a major pathway of iron uptake and translocation in rice. *J. Exp. Bot*. **72:** 2196–2211.
- Kong D, SA Khan, H Wu, Y Liu and Ling HQ (2022) Biofortification of iron and zinc in rice and wheat. *J Integr Plant Biol.* **64(6):** 1157-67.
- Lu L, S Tian, H Liao, J Zhang and X Yang (2013) Analysis of Metal Element Distributions in Rice (*Oryza sativa* L.) Seeds and Relocation during Germination Based on X-Ray Fluorescence Imaging of Zn, Fe, K, Ca, and Mn. *PLoS One*. **8(2):** e57360.
- Maret W and HH Sandstead (2006) Zinc requirements and the risks and benefits of zinc supplementation. *J. Trace Elem. Med. Biol.* **20(1):** 3-18.
- Nachimuthu VV, S Robin, D Sudhakar, S Rajeswari, M Raveendran, K Subramanian, S Tannidi, BA Pandian (2014) Genotypic Variation for Micronutrient Content in Traditional and Improved Rice Lines and its Role in Biofortification Programme Indian *J. Sci. Tech*. **7(9):** 1414–1425.
- Oikeh SO, A Menkir, BM Dixon, RM Welch, RP Glahn and G Gauch (2004) Environmental stability of iron and zinc concentrations in grain of elite early maturing tropical maize genotypes grown under field conditions. *J. Agric. Sci*. **142:** 543–51.
- Olsen LI and MG Palmgren (2014) Many rivers to cross: the journey of zinc from soil toseed. Front. Plant Sci. 5: 30.
- Omary MB, C Fong, J Rothschild and P Finney (2012) Effects of germination on thenutritional profile of gluten-free cereals and pseudo cereals: a review. *Cereal Chem.* **89(1):** 1-14.
- Parikh M, AK Sarawgi, D Rao and B Sharma (2019) Assessment of genotypic variability for grain zinc and iron content in traditional and improved rice genotypes using energy

dispersive X-ray fluorescence spectrophotometer (ED-XRF). *Int. J. Curr. Microbiol. App. Sci.* **7(1):** 1967-1974.

- Prasanna BM, S Mazumdar, M Chakraborti, F Hossain, KM Manjaiah, PK Agrawal, SK Guleria and HS Gupta (2011) Genetic variability and genotype × Environment interactions for kernel Iron and Zinc concentrations in maize (*Zea mays*) genotypes. *Indian J. Agric. Sci*. **81(8):** 704–11.
- Pyo E, BL Tsang and ME Parker (2022) Rice as a vehicle for micronutrient fortification: A systematic review of micronutrient retention, organoleptic properties, and consumer acceptability. *Nutr. Rev*. **80:** 1062–1085.
- Rao DS, PM Babu, P Swarnalatha, S Kota, VP Bhadana, GS Varaprasad and VR Babu (2014) Assessment of grain zinc and iron variability in rice germplasm using Energy Dispersive X-ray Fluorescence Spectrophotometer. *J. Rice Res*. **7(1):** 45-52.
- Roy SC and BD Sharma (2014) Assessment of genetic diversity in rice (*Oryza sativa* L.) germplasm based on agro-morphology traits and zinc-iron content for crop improvement. *Physiol. Mol. Biol. Plants,* **20(2):** 209–24.
- Ryu MS and TB Aydemir (2020) Zinc. In: *Present Knowledge in Nutrition (Eleventh Edition)*. BP Marriott, DF Birt, VA Stallings, AA Yates (Ed.), Academic Press, USA, p 393–408.
- Sanjeeva RD, CN Neeraja, BP Madhu, B Nirmala, K Suman, LV Rao, K Surekha, P Raghu, T Longvah, P Surendra and R Kumar (2020) Zinc biofortified rice varieties: challenges, possibilities, and progress in India. *Front. Nutr*. **7:** 26.
- Sellappan K, K Datta, V Parkhi and SK Datta (2009) Rice caryopsis structure in relation to distribution of micronutrients (iron,

zinc, b-carotene) of rice cultivars including transgenic indica rice. *Plant Sci.* **177:** 557–562.

- Shahzad Z, H Rouached and A Rakha (2014) Combating mineral malnutrition through iron and zinc biofortification of cereals. *Compr. Rev. Food Sci. Food Saf.* **13(3):** 329-346.
- Su D, F Sultan, NC Zhao, BT Lei, FB Wang, G Pan and FM Cheng (2014) Positional variation in grain mineral nutrients within a rice panicle and its relation to phytic acid concentration. *J. Zhejiang Univ. Sci. B.* **15(11):** 986-996.
- Suman K, CN Neeraja, P Madhubabu, S Rathod, S Bej, KP Jadhav, JA Kumar, U Chaitanya, SC Pawar, Rani SH, LV Subbarao and SR Voleti (2021) Identification of promising RILs for high grain zinc through genotype x environment analysis and stable grain zinc QTL using SSRs and SNPs in rice (Oryza sativa L.). *Front. Plant Sci*. **12:** 587482.
- Suwarto N (2011) Genotype×Environment Interaction for Iron Concentration of Rice in Central Java of Indonesia. *Rice Sci*. **18(1):** 75−78.
- Velu G, RP Singh, J Huerta-Espino, RJ Pena, B Arun, MA Singh, YM Mujahide, VS Sohuf, GS Mavif, J Crossaa, G Alvarado, AK Joshi and WH Pfeiffer (2012) Performance of biofortified spring wheat genotypes in target environments for grain zinc and iron concentrations. *Field Crops Res.* **137:** 261–267.
- Zhang GM, TQ Zheng, Z Chen, YL Wang, Y Wang, YM Shi, CC Wang, LY Zhang, JT Ma, LW Deng, W Li, TT Xu, CZ Liang, JL Xu and ZK Li (2018) Joint exploration of favorable haplotypes for mineral concentrations in milled grains of rice (*Oryza sativa* L.). *Front. Plant Sci*. **9:** 447.