

Quarantine Risk Associated with Exchange of Plant Genera Carrying Hidden Infestation

Shashi Bhalla, Manju Lata Kapur, B Lal, BR Verma and Charan Singh

Division of Plant Quarantine, National Bureau of Plant Genetic Resources, New Delhi-110 012

Key Words: Latent/Hidden Infestation, Plant Genera, Quarantine, X-ray Radiography

A large number of plant germplasm samples are being exchanged throughout the world for crop improvement programmes. In addition, the liberalized trade in the present global scenario (WTO regime) has increased the flow of agricultural commodities across different countries. These exchanges always carry the risk of inadvertent introduction and spread of exotic pests. Such pests, if established may prove more devastating in the new geographical areas in the absence of natural enemies and competitors. The risk of introduction of exotic pests could be minimized by undertaking quarantine of the planting material under exchange. The effective implementation of quarantine measures at National Bureau of Plant Genetic Resources (NBPGR) has resulted in the interception of a number of exotic pests. These include *Acanthoscelides obtectus* in *Cajanus cajan*, *Phaseolus vulgaris* and *Vicia faba* from Brazil, Bulgaria and Malawi; *Anthonomus grandis* in *Gossypium* sp. and *Hibiscus* spp. from USA; *Bruchophagus gibbus* in *Medicago* spp. and *Trifolium* spp. from France, Germany, Italy, USA and USSR; *Leptinotarsa decemlineata* in potato from Europe and USA; *Popillia japonica* in root stocks of nursery crops from Japan, USA and USSR.

There are groups of insects infesting seeds, viz., seed wasps (Hymenoptera: Chalcidoidea), pulse beetles (Coleoptera: Bruchidae), and a few weevils/borers (Coleoptera: Curculionidae/Scolytidae) which infest the developing seeds without showing external symptoms of their presence within. Such an infestation is called hidden or latent infestation.

The quarantine risk becomes much higher with the exchange of seeds with hidden infestation because these are not detectable through routine visual inspection. There are several examples of movement of such insects across the geographical boundaries. The douglas fir seed chalcid (*Megastigmus spermotrophus* Wachtl.) has spread from its original home in western United States to Scotland (Mac Dougall, 1926), France (Vayssiere, 1931), Germany (Escherich, 1938) and New Zealand (courlay, 1930)

alongwith the shipment of douglas fir (*Pseudotsuga taxifolia*) seeds. The rose seed chalcid [*Megastigmus aculeatus* (Swedrus)] was introduced into New Jersey (USA) through the seeds of *Rosa multiflora* from Japan (Weiss, 1917). The crop losses reported to have been caused by various introduced chalcid pests include seed damage over 80% by *Bruchophagus roddi* in the seeds of alfalfa (Strong, 1962) and about 40% by *Systole albipennis* in various umbelliferous seeds (Gupta, 1962). Seed damage to the extent of about 40% has been reported in *Trifolium alexandrinum* by bruchids, viz., *Bruchidius alfieri* and *Bruchidius trifolii* (Abou-Raya, 1954).

Detection of Hidden Infestation

During quarantine processing, the seeds of *Glycyrrhiza glabra* imported from Iraq were suspected to carry hidden infestation. Hence, these were detained to observe the emergence of any insect. The insects emerged during detention were identified as eurytomids (Chalcidoidea), *Bruchophagus glycyrrhizae* (Wadhi, 1963, 1967) and bruchid, *Bruchidius* sp. (Wadhi *et al.*, 1966). This highlighted the possibility of escape of such pests during routine quarantine inspection due to hidden nature of infestation and the need for development of specialized detection procedures. Therefore, X-ray radiography technique was standardized for the detection of hidden infestation in seeds at National Bureau of Plant Genetic Resources (Wadhi *et al.*, 1967). The seeds are spread on a sheet of paper that is placed on an envelope containing the Agfa Rapid X-ray film and are exposed to soft X-rays generated at 22 Kv, 3 mA, at a distance of 30 cm from the source for 15 seconds. The envelope containing the exposed film is removed gently without disturbing the seed geometry and the film is developed/fixer prepared for the purpose. Infested seeds are marked on the X-ray plate and corresponding (infested) seeds are then hand picked from the seed sample with original geometry retained on the paper. The dose, however, is varied with the size, shape or thickness of the seeds

for better resolution of image on the X-ray plate. The dose is also standardized for different seeds as and when required, as was done in case of mango stones (exceptionally large seeds) for detection of mango stone weevil, *Sternochaetus mangiferae* (Kapur *et al.*, 1997).

The seed transparency technique (Kaura, 1959) is used for detection of hidden infestation in small seeds as it is difficult to screen them through X-ray radiography. The seeds are boiled in lactophenol solution containing distilled water, phenol crystals, lactic acid and glycerine till these are sufficiently transparent to reveal the internal infestation. However, the fumes of phenol (skin irritant) are hazardous to the workers and also the valuable imported material under test gets destroyed during the process. These two techniques have been extensively used to screen the seeds of plant genera carrying hidden infestation since 1967 (Wadhi *et al.*, 1968).

Techniques such as X-ray micrography or projection micrography need to be developed so that the radiograph of small seeds could be enlarged and accurately analysed. This would overcome the limitations of two techniques.

Need to Identify the Plant Genera Carrying Hidden Infestation

X-ray radiography is a cumbersome and time consuming process. It also requires enormous funds and, thus, it is practically impossible to screen all the exotic seed material. Therefore, need was felt to identify the plant genera which can carry the latent infestation so as to reduce the workload of X-ray radiography and to minimize the quarantine risk of introduction of pests causing latent infestation. Consequently, two lists namely one for plant genera reported to be infested by one or the other species of *Phytophagous chacidoids* (Wadhi, 1967) and other for bruchids of the genus *Bruchidius* (Wadhi and Verma, 1971) were drawn from the published reports. These two lists comprised 148 plant genera that were designated as the *listed plant genera* and their seeds were compulsorily subjected to X-ray radiography (Wadhi, 1980).

Later, during the quarantine processing, seeds of genera other than the listed plant genera were also suspected to carry infestation and the same was confirmed through X-ray screening. The bruchid, *Pachymeus lacerrdae* (Coleoptera: Bruchidae) was intercepted in the nuts of *Orbignya phalerata* from Brazil (Kapur *et al.*, 1988); seed chalcid, *Eurytoma* sp. in the seeds of *Urginea maritima* from Cyprus (Kapur *et al.*, 1992); seed chalcids, *Quadrastichodella eucalypti* in seeds of

Eucalyptus and *Bootanomyia* sp. In seeds of *Casurina* sp. from Australia (Mathur and Lal, 1996). Analysis of pest risk in respect of various crops through perusal of literature (Kapur *et al.*, 1989) and over experience of exposing the doubtful seeds to X-ray radiography resulted in addition of many more genera to the listed plant genera, requiring X-ray screening. The list was further updated by extensive scrutiny of the catalogues on family bruchidae (Udayagiri and Wadhi, 1989) and phytophagous chalcidoidea (Muesebeck *et al.*, 1975; Krombein, 1958; Peck, 1963), review of Agricultural Entomology, Series A (Anonymous, 2000) and available literature. The list now comprises 340 plant genera belonging to 75 plant families. The families of plant genera (Jackson, 1977a,b; Usher, 1984; Terrell *et al.*, 1986) have been included for future reference because it is well known that the host plant relationship exists between specific taxonomic groups of plants and insects. This would help as a precaution against other plant genera also belonging to these families which may harbour similar infestation. The updated list (Table 1) shows plant genera known to carry hidden infestation of bruchids (194), chalcids (72), both bruchids and chalcids (71) and others as curculionids/scolytids (3).

Table 1. Plant genera in their seeds which carry hidden infestation of bruchids, chalcids and others

S.No.	Genus	Family	S.No.	Genus	Family
1.	<i>Abelmoschus</i> ^b	Malvaceae	33.	<i>Astrocaryum</i> ^b	Palmae
2.	<i>Abies</i> ^c	Pinaceae	34.	<i>Atalea</i> ^b	Araceae
3.	<i>Abrus</i> ^b	Fabaceae	35.	<i>Arylosia</i> ^b	Fabaceae
4.	<i>Abutilon</i> ^b	Malvaceae	36.	<i>Baccharis</i> ^b	Compositae
5.	<i>Acacia</i> ^b	Fabaceae	37.	<i>Bactris</i> ^b	Arecaceae
6.	<i>Acer</i> ^c	Aceraceae	38.	<i>Bambusa</i> ^c	Poaceae
7.	<i>Achillea</i> ^c	Asteraceae	39.	<i>Banisteriopsis</i> ^b	Malpighiaceae
8.	<i>Acrocomia</i> ^b	Arecaceae	40.	<i>Baptisia</i> ^b	Fabaceae
9.	<i>Acauan</i> ^b	Fabaceae	41.	<i>Barcena</i> ^b	Fabaceae
10.	<i>Adenocarpus</i> ^b	Fabaceae	42.	<i>Bauhinia</i> ^b	Fabaceae
11.	<i>Aeschynomene</i> ^b	Fabaceae	43.	<i>Bellis</i> ^c	Compositae
12.	<i>Albizia</i> ^b	Fabaceae	44.	<i>Bixa</i> ^b	Bixaceae
13.	<i>Alhagi</i> ^b	Fabaceae	45.	<i>Bonjeana</i> ^c	Fabaceae
14.	<i>Allocauarina</i> ^c	Casuarinaceae	46.	<i>Borago</i> ^b	Boraginaceae
15.	<i>Aloe</i> ^c	Aloeaceae	47.	<i>Bothriochloa</i> ^c	Poaceae
16.	<i>Amelanchier</i> ^c	Rosaceae	48.	<i>Bradburia</i> ^b	Compositae
17.	<i>Ammi</i> ^c	Apiaceae	49.	<i>Brassica</i> ^b	Brassicaceae
18.	<i>Ammodendron</i> ^b	Fabaceae	50.	<i>Bromus</i> ^c	Poaceae
19.	<i>Amomum</i> ^b	Zingiberaceae	51.	<i>Buchenania</i> ^b	Anacardiaceae
20.	<i>Amorpha</i> ^b	Fabaceae	52.	<i>Bupleurum</i> ^c	Apiaceae
21.	<i>Anethum</i> ^c	Apiaceae	53.	<i>Butea</i> ^b	Fabaceae
22.	<i>Annona</i> ^c	Annonaceae	54.	<i>Cachrys</i> ^c	Apiaceae
23.	<i>Anthyllis</i> ^c	Fabaceae	55.	<i>Caesalpinia</i> ^b	Fabaceae
24.	<i>Apeiba</i> ^b	Tiliaceae	56.	<i>Cajanus</i> ^b	Fabaceae
25.	<i>Apium</i> ^c	Apiaceae	57.	<i>Calapogonium</i> ^b	Fabaceae
26.	<i>Apoplanesia</i> ^b	Fabaceae	58.	<i>Calliandra</i> ^b	Fabaceae
27.	<i>Arachis</i> ^b	Fabaceae	59.	<i>Calycotome</i> ^c	Fabaceae
28.	<i>Arenaria</i> ^b	Caryophyllaceae	60.	<i>Calystegia</i> ^b	Convolvulaceae
29.	<i>Argyreia</i> ^b	Convolvulaceae	61.	<i>Camellia</i> ^c	Theaceae
30.	<i>Artemisia</i> ^b	Asteraceae	62.	<i>Canavalia</i> ^b	Fabaceae
31.	<i>Asclepias</i> ^c	Asclepiadaceae	63.	<i>Capparis</i> ^b	Capparaceae
32.	<i>Astragalus</i> ^c	Fabaceae	64.	<i>Caragana</i> ^c	Fabaceae

S.No. Genus	Family	S.No. Genus	Family	S.No. Genus	Family	S.No. Genus	Family
65. <i>Cassia</i> ^b	Fabaceae	136. <i>Erythrina</i> ^b	Fabaceae	207. <i>Medicago</i> ^{bc}	Fabaceae	279. <i>Ricinus</i> ^b	Euphorbiaceae
66. <i>Casuarina</i> ^b	Casuarianaceae	137. <i>Eucalyptus</i> ^c	Myrtaceae	208. <i>Meibomia</i> ^b	Fabaceae	280. <i>Rhinia</i> ^{bc}	Fabaceae
67. <i>Cucalis</i> ^{bc}	Apiaceae	138. <i>Eugenia</i> ^{bc}	Myrtaceae	209. <i>Melanoxylon</i> ^b	Fabaceae	281. <i>Rosa</i> ^{bc}	Rosaceae
68. <i>Cedrus</i> ^c	Pinaceae	139. <i>Eupatorium</i> ^b	Asteraceae	210. <i>Melilotus</i> ^{bc}	Fabaceae	282. <i>Rosmarinus</i> ^c	Lamiaceae
69. <i>Cenchrus</i> ^c	Poaceae	140. <i>Euphorbia</i> ^{bc}	Euphorbiaceae	211. <i>Mentha</i> ^{bc}	Lamiaceae	283. <i>Ruellia</i> ^b	Acanthaceae
70. <i>Centaura</i> ^{bc}	Asteraceae	141. <i>Euterpe</i> ^b	Arecaceae	212. <i>Merremia</i> ^b	Convolvulaceae	284. <i>Saba</i> ^b	Arecaceae
71. <i>Centranthus</i> ^b	Velarianaeae	142. <i>Fagopyrum</i> ^b	Polygonaceae	213. <i>Mimosa</i> ^b	Fabaceae	285. <i>Salvia</i> ^{bc}	Lamiaceae
72. <i>Centrosema</i> ^b	Fabaceae	143. <i>Festuca</i> ^c	Poaceae	214. <i>Mucuna</i> ^b	Fabaceae	286. <i>Samanea</i> ^{bc}	Fabaceae
73. <i>Cerantonia</i> ^b	Fabaceae	144. <i>Ficus</i> ^c	Moraceae	215. <i>Myobroma</i> ^b	Fabaceae	287. <i>Sarothamnus</i> ^b	Fabaceae
74. <i>Cercidium</i> ^b	Fabaceae	145. <i>Foeniculum</i> ^f	Apiaceae	216. <i>Myrcia</i> ^b	Myrtaceae	288. <i>Sauvagesia</i> ^b	Ochnaceae
75. <i>Cercis</i> ^b	Fabaceae	146. <i>Galactia</i> ^b	Fabaceae	217. <i>Nelumbo</i> ^b	Nelumbonaceae	289. <i>Scheelea</i> ^a	Plamae
76. <i>Chamaecrista</i> ^b	Fabaceae	147. <i>Galega</i> ^{bc}	Fabaceae	218. <i>Neptunia</i> ^b	Fabaceae	290. <i>Schinus</i> ^a	Anacardiaceae
77. <i>Chamaecruistata</i> ^c	Pinaceae	148. <i>Genista</i> ^{bc}	Fabaceae	219. <i>Nissolia</i> ^b	Fabaceae	291. <i>Schrunkia</i> ^b	Fabaceae
78. <i>Chaemaecyparis</i> ^b	Cupressaceae	149. <i>Gleditschia</i> ^{bc}	Fabaceae	220. <i>Nitraria</i> ^b	Zygophyllaceae	292. <i>Scorpiurus</i> ^{bc}	Fabaceae
79. <i>Chesneya</i> ^b	Fabaceae	150. <i>Glycine</i> ^{bc}	Fabaceae	221. <i>Ochroma</i> ^b	Bombacaceae	293. <i>Senna</i> ^b	Febacea
80. <i>Chrysanthemum</i> ^{bc}	Asteraceae	151. <i>Glycyrrhiza</i> ^{bc}	Fabaceae	222. <i>Olea</i> ^c	Oleaceae	294. <i>Sesamum</i> ^f	Pedaliaceae
81. <i>Cicer</i> ^b	Fabaceae	152. <i>Gossypium</i> ^{bc}	Malvaceae	223. <i>Olneya</i> ^b	Fabaceae	295. <i>Sesbania</i> ^{bc}	Fabaceae
82. <i>Cichorium</i> ^f	Asteraceae	153. <i>Guarea</i> ^b	Meliaceae	224. <i>Onobrychis</i> ^{bc}	Fabaceae	296. <i>Sesbastiana</i> ^b	Euphorbiaceae
83. <i>Cissus</i> ^{bc}	Vitaceae	154. <i>Guazuma</i> ^b	Sterculiaceae	225. <i>Ononis</i> ^{bc}	Fabaceae	297. <i>Sideroxylon</i> ^b	Sapotaceae
84. <i>Cnidocolus</i> ^c	Euphorbiaceae	155. <i>Gymnosperma</i> ^b	Compositae	226. <i>Orbignya</i> ^b	Arecaceae	298. <i>Silybum</i> ^f	Asteraceae
85. <i>Coccolrinax</i> ^c	Arecaceae	156. <i>Haematoxylon</i> ^f	Fabaceae	227. <i>Ormithopus</i> ^{bc}	Fabaceae	299. <i>Smilax</i> ^c	Smilacaceae
86. <i>Cocos</i> ^b	Arecaceae	157. <i>Halimodendron</i> ^{bc}	Fabaceae	228. <i>Oryza</i> ^b	Poaceae	300. <i>Smirnowia</i> ^c	Fabaceae
87. <i>Colutea</i> ^b	Fabaceae	158. <i>Hardwickia</i> ^b	Fabaceae	229. <i>Oxytropis</i> ^{bc}	Fabaceae	301. <i>Soja</i> ^b	Fabaceae
88. <i>Combretum</i> ^{bc}	Combretaceae	159. <i>Hedysarum</i> ^{bc}	Fabaceae	230. <i>Pachyrrhizus</i> ^b	Fabaceae	302. <i>Sophora</i> ^{bc}	Fabaceae
89. <i>Condalia</i> ^b	Rhamnaceae	160. <i>Helianthemum</i> ^b	Cistaceae	231. <i>Panax</i> ^c	Araliaceae	303. <i>Sorbus</i> ^c	Rosaceae
90. <i>Convolvulus</i> ^b	Arecaceae	161. <i>Helianthus</i> ^b	Asteraceae	232. <i>Pandanus</i> ^b	Pandanaceae	304. <i>Spartium</i> ^{bc}	Fabaceae
91. <i>Copernicia</i> ^b	Arecaceae	162. <i>Heracleum</i> ^b	Umbelliferae	233. <i>Panicum</i> ^b	Poaceae	305. <i>Sphaerophysa</i> ^b	Fabaceae
92. <i>Cordia</i> ^b	Boraginaceae	163. <i>Heteropterys</i> ^b	Malpighiaceae	234. <i>Parkia</i> ^b	Fabaceae	306. <i>Spiraea</i> ^c	Rosaceae
93. <i>Coriandrum</i> ^c	Apiaceae	164. <i>Hibiscus</i> ^c	Malvaceae	235. <i>Parkinsonia</i> ^{bc}	Fabaceae	307. <i>Spondias</i> ^{bc}	Anacardiaceae
94. <i>Coronilla</i> ^{bc}	Fabaceae	165. <i>Hippocrepis</i> ^{bc}	Fabaceae	236. <i>Parryella</i> ^b	Fabaceae	308. <i>Strombocarpus</i> ^b	Fabaceae
95. <i>Corsotia</i> ^b	Fabaceae	166. <i>Hippomane</i> ^b	Euphorbiaceae	237. <i>Parthenium</i> ^b	Asteraceae	309. <i>Strophostyles</i> ^{bc}	Fabaceae
96. <i>Cracca</i> ^b	Fabaceae	167. <i>Hoffmanseggia</i> ^b	Fabaceae	238. <i>Parthenocissus</i> ^c	Vitaceae	310. <i>Stylosanthes</i> ^b	Fabaceae
97. <i>Crataegus</i> ^c	Rosaceae	168. <i>Holcus</i> ^c	Poaceae	239. <i>Pavonia</i> ^c	Malvaceae	311. <i>Tachigalia</i> ^b	Fabaceae
98. <i>Crotalaria</i> ^b	Fabaceae	169. <i>Hosackia</i> ^b	Fabaceae	240. <i>Pennisetum</i> ^b	Poaceae	312. <i>Tamarindus</i> ^b	Fabaceae
99. <i>Crotomeria</i> ^c	Coniferaceae	170. <i>Hymenaea</i> ^b	Fabaceae	241. <i>Petalostemon</i> ^b	Fabaceae	313. <i>Tamarix</i> ^c	Tamaricaceae
100. <i>Cumin</i> ^c	Apiaceae	171. <i>Hymenoclea</i> ^b	Compositae	242. <i>Petit</i> ^f	Verbenaceae	314. <i>Taverniera</i> ^b	Fabaceae
101. <i>Cupressus</i> ^{bc}	Cupressaceae	172. <i>Isma</i> ^b	Compositae	243. <i>Petturia</i> ^{bc}	Fabaceae	315. <i>Tectona</i> ^b	Verbenaceae
102. <i>Cyamopsis</i> ^{bc}	Fabaceae	173. <i>Ichthyomethia</i> ^b	Fabaceae	244. <i>Phaseolus</i> ^b	Fabaceae	316. <i>Tephrosia</i> ^{bc}	Fabaceae
103. <i>Cyrtistax</i> ^b	Bignoniaceae	174. <i>Ilex</i> ^c	Aquifoliaceae	245. <i>Phleum</i> ^f	Poaceae	317. <i>Terminalia</i> ^b	Combretaceae
104. <i>Cymbosema</i> ^b	Fabaceae	175. <i>Indigofera</i> ^c	Fabaceae	246. <i>Phlox</i> ^c	Arecaceae	318. <i>Tetragolobus</i> ^b	Fabaceae
105. <i>Cynoglossum</i> ^f	Boraginaceae	176. <i>Inodes</i> ^b	Palmae	247. <i>Physocarpus</i> ^c	Rosaceae	319. <i>Theobroma</i> ^b	Sterculiaceae
106. <i>Cytisus</i> ^b	Fabaceae	177. <i>Inula</i> ^c	Asteraceae	248. <i>Physostigma</i> ^b	Fabaceae	320. <i>Thermopsis</i> ^{bc}	Fabaceae
107. <i>Dactylis</i> ^c	Poaceae	178. <i>Impmoeda</i> ^b	Convolvulaceae	249. <i>Phytelphas</i> ^b	Arecaceae	321. <i>Thyslopsis</i> ^b	Coniferaceae
108. <i>Dahlia</i> ^c	Compositae	179. <i>Isatis</i> ^c	Brassicaceae	250. <i>Picea</i> ^c	Pinaceae	322. <i>Tillandsia</i> ^b	Bromeliaceae
109. <i>Dalbergia</i> ^b	Fabaceae	180. <i>Juniperus</i> ^c	Cupressaceae	251. <i>Piliostigma</i> ^b	Fabaceae	323. <i>Trifolium</i> ^{bc}	Fabaceae
110. <i>Dalea</i> ^b	Fabaceae	181. <i>Lab lab</i> ^b	Fabaceae	252. <i>Pimpinella</i> ^b	Apiaceae	324. <i>Trigonella</i> ^{bc}	Fabaceae
111. <i>Daubentonia</i> ^b	Fabaceae	182. <i>Laburnum</i> ^{bc}	Fabaceae	253. <i>Pinus</i> ^c	Pinaceae	325. <i>Triumfetta</i> ^b	Tiliaceae
112. <i>Daucus</i> ^{bc}	Apiaceae	183. <i>Lagonychium</i> ^b	Fabaceae	254. <i>Piptadenia</i> ^b	Fabaceae	326. <i>Tsuga</i> ^c	Pinaceae
113. <i>Dendrocalamus</i> ^c	Poaceae	184. <i>Larix</i> ^c	Pinaceae	255. <i>Piscidia</i> ^b	Fabaceae	327. <i>Ulex</i> ^c	Fabaceae
114. <i>Desmanthus</i> ^b	Fabaceae	185. <i>Laserpitium</i> ^f	Umbelliferae	256. <i>Pistacia</i> ^c	Anacardiaceae	328. <i>Ulmus</i> ^c	Ulmaceae
115. <i>Desmodium</i> ^{bc}	Fabaceae	186. <i>Lathyrus</i> ^{bc}	Fabaceae	257. <i>Pisum</i> ^b	Fabaceae	329. <i>Urena</i> ^b	Malvaceae
116. <i>Desmonchus</i> ^b	Plamae	187. <i>Lechea</i> ^b	Cistaceae	258. <i>Pithecellobium</i> ^b	Fabaceae	330. <i>Urginea</i> ^b	Liliaceae
117. <i>Dialium</i> ^b	Fabaceae	188. <i>Lens</i> ^{bc}	Fabaceae	259. <i>Plantago</i> ^b	Plantaginaceae	331. <i>Vachellia</i> ^b	Mimosaseae
118. <i>Dichostachys</i> ^{bc}	Fabaceae	189. <i>Lepidium</i> ^f	Brassicaceae	260. <i>Poinciana</i> ^b	Fabaceae	332. <i>Verbascum</i> ^c	Scrophylariaceae
119. <i>Dioclea</i> ^b	Fabaceae	190. <i>Leptoglottis</i> ^b	Fabaceae	261. <i>Podocytisus</i> ^{bc}	Fabaceae	333. <i>Veronica</i> ^c	Scrophylariaceae
120. <i>Diospyros</i> ^{bc}	Ebenaceae	191. <i>Lepedeza</i> ^{bc}	Fabaceae	262. <i>Prangos</i> ^{bc}	Umbelliferae	334. <i>Viburnum</i> ^c	Caprifoliaceae
121. <i>Dodonaea</i> ^b	Sapindaceae	192. <i>Leuceana</i> ^b	Fabaceae	263. <i>Primula</i> ^c	Primulaceae	335. <i>Vicia</i> ^b	Fabaceae
122. <i>Dolichos</i> ^{bc}	Fabaceae	193. <i>Leuzea</i> ^b	Compositae	264. <i>Prosopis</i> ^{bc}	Fabaceae	336. <i>Vigna</i> ^b	Fabaceae
123. <i>Dombeya</i> ^b	Sterculiaceae	194. <i>Lisaeda</i> ^b	Umbelliferae	265. <i>Prunus</i> ^c	Rosaceae	337. <i>Vitis</i> ^{bc}	Vitaceae
124. <i>Dorycnium</i> ^b	Fabaceae	195. <i>Lithraea</i> ^b	Anacardiaceae	266. <i>Pseudabutylon</i> ^b	Malvaceae	338. <i>Voandzeia</i> ^b	Fabaceae
125. <i>Draba</i> ^c	Cruciferae	196. <i>Lonchocarpus</i> ^b	Fabaceae	267. <i>Pseudosamanea</i> ^b	Fabaceae	339. <i>Zea</i> ^b	Poaceae
126. <i>Drypetes</i> ^b	Euphorbiaceae	197. <i>Lotus</i> ^{bc}	Fabaceae	268. <i>Pseudotsuga</i> ^c	Pinaceae	340. <i>Zizyphus</i> ^b	Rhamnaceae
127. <i>Earleocassia</i> ^b	Fabaceae	198. <i>Luchia</i> ^b	Tiliaceae	269. <i>Psidium</i> ^f	Myrtaceae		
128. <i>Echium</i> ^c	Boraginaceae	199. <i>Ludwigia</i> ^b	Onagraceae	270. <i>Psophocarpus</i> ^b	Fabaceae		
129. <i>Elaeis</i> ^b	Arecaceae	200. <i>Lupinus</i> ^{bc}	Fabaceae	271. <i>Pterocarpus</i> ^b	Fabaceae		
130. <i>Enterolobium</i> ^b	Fabaceae	201. <i>Lysiloma</i> ^b	Fabaceae	272. <i>Pteraria</i> ^b	Fabaceae		
131. <i>Epilobium</i> ^b	Onagraceae	202. <i>Lythrum</i> ^b	Lythraceae	273. <i>Pyrus</i> ^c	Rosaceae		
132. <i>Eriosema</i> ^b	Fabaceae	203. <i>Macroptilium</i> ^b	Fabaceae	274. <i>Quercus</i> ^c	Fagaceae		
133. <i>Erazurizia</i> ^b	Fabaceae	204. <i>Malus</i> ^c	Rosaceae	275. <i>Roseda</i> ^c	Resedaceae		
134. <i>Eryngium</i> ^{bc}	Apiaceae	205. <i>Mangifera</i> ^b	Anacardiaceae	276. <i>Rhamnus</i> ^b	Rhamnaceae		
135. <i>Erythraea</i> ^b	Gentianaceae	206. <i>Mecadamia</i> ^b	Proteaceae	277. <i>Rhus</i> ^c	Anacardiaceae		
				278. <i>Rhynchosia</i> ^b	Fabaceae		

b = Plant genera known to carry hidden infestation of Bruchids

c = Plant genera known to carry hidden infestation of Chalcids

bc = Plant genera known to carry hidden infestation of Bruchids and Chalcids

0 = Plant genera known to carry hidden infestation of other (Curculionids/ Scolytids etc.)

Several interceptions of exotic bruchids, chalcids etc. of quarantine significance have been made in the Plant Quarantine Division as a result of X-ray screening of the listed plant genera (Verma, 1980; Chandel and Singh, 1987; Mathur and Lal, 1996). This information is of paramount importance for the exchange of seeds including transgenics among different countries. Therefore, it is suggested to expose the listed plant genera to X-ray radiography at all the ports of entry.

Acknowledgements

The authors are grateful to the Head, Division of Plant Quarantine for continuous support, guidance and encouragement and to the Director, NBPGR, New Delhi, for providing the facilities.

References

- Anonymous (2000) *Review of Agricultural Entomology Series A*, CAB, UK.
- Abou-Raya AK (1954) *Bruchidius alferi* Pic a biologic race of *Bruchidius trifolii* Motsch. (Coleoptera: Bruchidae). *Bull. Soc. Fouad I. Ent.* **38**: 193-203.
- Chandel KPS and BM Singh (1987) *Research Achievements-A Decade. NBPGR Sci. Monogr. No. 11*, 170p.
- Escherich K (1938) The phytophagous species of *Megastigmus* as destroyer of the seeds of conifers. *Rev. Appl. Ent. (A)* **27**: 230.
- Gourlay ES (1930) Some parasitic hymenoptera of economic importance in New Zealand. *N.Z.J. Sci. Tech.* **11**: 319.
- Gupta SC (1962) Occurrence of exembryonate seeds in the umbelliferae. *Curr. Sci.* **31**: 203-205.
- Jackson BD (1977a) *Index Kewensis-An Enumeration of the Genera and Species of Flowering Plants*. Otto Koeltz Science Publishers, West Germany. **1**: 1268p.
- Jackson BD (1977b) *Index Kewensis-An Enumeration of the Genera and Species of Flowering Plants*. Otto Koeltz Science Publishers, West Germany. **2**: 1300p.
- Kapur Manju Lata, BR Verma and B Lal (1988) Interception of *Pachymerus lacerdae* (Coleoptera: Bruchidae) from imported nuts of *Orbignya phalerata* (Palmae)—a new host record. *J. Young Ent. Quart.* **6**: 31-32.
- Kapur Manju Lata, B Lal and BR Verma (1989) Studies on pest risk involved in import of medicinal plants. *Indian J. Pl. Genet. Resour.* **2**: 55-59.
- Kapur Manju Lata, B Lal and BR Verma (1992) *Eurytoma* intercepted from imported seeds of *Urginea maritima*—A first record. *Indian J. Ent.* **55**: 451-52.
- Kapur Manju Lata, BR Verma and Shashi Bhalla (1997) Quarantine detection of mango stone weevil, *Sternochaetus mangifera* (F.) in mango through X-ray radiography. In: *Souvenir and Abstracts: Symposium on Integrated Pest Management for Sustainable Crop Production*, IARI, New Delhi, pp. 59.
- Kaura (1959) A new transparency method for detecting internal infestation in grains. *Grain Storage Newslet.* **1**: 12.
- Krombein Karl V (1958) *Hymenoptera of America North of Mexico. USDA Agriculture Monogr. 2*: 305p.
- Mac Dougall RS (1926) The *Megastigmus* of the douglas fir seed. *Trans. R. Scot. Arbor. Soc.* **19**: 52-65.
- Mathur VK and A Lal (1996) *Plant Quarantine Activities at NBPGR (1976-1996). NBPGR Sci. Monogr. No. 6*: 96p.
- Muesebeck CFW, Karl V Krombein and Henry K Townes (1951) *Hymenoptera of America North of Mexico. USDA*, 1420p.
- Peck O (1963) *A Catalogue of the Nearctic Chalcidoidea (Insecta: Hymenoptera)*. Canadian Entomologist Supplement No. 30, 1092p.
- Strong FE (1962) Laboratory studies on the biology of the alfalfa seed chalcid, *Bruchophagus roddi* Guss. (Hymenoptera: Eurytomidae). *Hilgardia* **32**: 229-249.
- Terrell EE, SR Hill, JH Wierseman and WE Rice (1986) *A Checklist of the Name of 3,000 Vascular Plants of Economic Importance. USDA, ARS, Agriculture Handbook*, 505p.
- Udayagiri S and SR Wadhi (1989) *Catalogue of Bruchidae. Mem. Amer. Ent. Inst. No. 45*, 300p.
- Usher G (1984) *A Dictionary of Plants Used by Man*. CBS Publishers & Distributors 620p.
- Vayssiere P (1931) Appartition en France du *Megastigmus spermotrophus* Wachtl. parasite des semences du *Pseudotsuga douglasii* Harr. *Rev. Appl. Ent. (A)* **19**: 586.
- Verma BR (1980) Pests intercepted from planting material imported during the period from January, 1956 to December, 1979. In: SR Wadhi (ed) *Plant Quarantine Activity at the National Bureau of Plant Genetic Resources. NBPGR Sci. Monograph No. 2*, pp 55-71.
- Wadhi SR (1963) Interception of *Bruchophagus glycyrrhizae* Nikolskaya from liquorice seeds. *Indian J. Ent.* **25**: 382-384.
- Wadhi SR (1967) Seed infesting chalcidoids and plant quarantine. *Indian J. Ent.* **29**: 252-260.
- Wadhi SR (1980) *Plant Quarantine Activity at the National Bureau of Plant Genetic Resources. NBPGR Sci. Monogr. 2*: p. 99.
- Wadhi SR and BR Verma (1971) Quarantine problem posed by *Bruchidius* species. *Indian J. Ent.* **33**: 34-41.
- Wadhi SR, BR Verma, Thomas Soares, Nirmolini Singh and Rattan Lal (1967) Detection of phytophagous chalcidoids in seeds for quarantine purpose. *Indian J. Ent.* **29**: 197-199.
- Wadhi SR, BR Verma, Thomas Soares, Nirmolini Singh and Rattan Lal (1968) Insects intercepted from the seed samples imported and exported for plant introduction work during January, 1965 to December, 1996. *Indian J. Ent.* **30**: 303-308.
- Wadhi SR, Thomas Soares and Rattan Lal (1966) Insect interceptions from the seed samples imported and exported for plant introduction work during the period from August, 1960 to December, 1964. *Indian J. Ent.* **28**: 494-500.
- Weiss HB (1917) *Megastigmus aculeatus* Swedrus-introduced into New Jersey from Japan. *J. Econ. Ent.* **6**: 402-403.