

## Comparative Performance of Sugarcane Genotypes against Stalk Borer (*Chilo auricilius* Dudgeon) in Punjab

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The present study was aimed at identification of the resistant/resistant clones of sugarcane against stalk borer (*Chilo auricilius*) for their exploitation in sugarcane economic breeding as the losses incurred by it to sugarcane cultivation in sub-tropical zone are upto 33% and there is no chemical control available for combating this pest in Punjab. The natural incidence of sugarcane stalk borer on 132 sugarcane genotypes/cultivars/elite clones/varieties, collected from 12 different sugarcane research centres of India, over two years has been reported. The experimental material exhibited large variations in the incidence of stalk borer ranging from 1 (Co Se 00421, Sel 18-01) to 32.91% (Co S 02258). Thirty-three genotypes showed less than five% incidence, categorized as resistant and, hence, suggested for genetic enhancement.

**Key Words:** Germplasm, Resistance, Stalk borer, Sugarcane

Amongst the large number of insect pests that regularly attack sugarcane crop, stalk borer (*Chilo auricilius* Dudgeon) is one of the most serious pests limiting sugarcane production especially in northern India, which can cause a loss upto 33% in cane yield and the 20.4% reduction in sucrose in juice (Jayanthi and Goud, 2002). It is active throughout the year but peak activity period is observed from July-February under Punjab conditions.

The losses caused by the borer must be minimized to increase profits but effective chemical control methods against this pest are not available and feasible (Chaudhary *et al.*, 1980). Aggravating this problem, the mechanical and cultural control methods against stalk borer are laborious and usually not followed by farmers. Hence, breeding for resistant varieties is the cheapest, reliable and most suitable method for the control of this pest. Keeping this consideration in view, the evaluation of genotypes for reaction against stalk borer was undertaken to identify promising genotypes.

### Materials and Methods

The field screening was conducted on 132 sugarcane genotypes/cultivars/elite clones/varieties, collected from 12 different sugarcane research centres of India under All India Coordinated Research Programme on Sugarcane (AICRP-S). These research centres included Indian Institute of Sugarcane Research, Lucknow, Uttar Pradesh (UP); Sugarcane Breeding Institute (SBI), Coimbatore, Tamil Nadu; Chaudhary Charan Singh Haryana Agricultural University (CCSHAU), Hisar, Haryana;

Punjab Agricultural University (PAU), Ludhiana, Punjab; GB Pant University of Agriculture and Technology (GBPUAT), Pant Nagar, Uttarakhand; Rajendra Agricultural University (RAU), Pusa, Bihar; Acharya NG Ranga Agricultural University (ANGRAU), Hyderabad, AP; Orissa University of Agriculture and Technology (OUAT), Bhubaneswar, Orissa; UP Council of Sugarcane research, Shahjahanpur, UP; GS Sugarcane breeding and research institute, Seorahi, UP, Regional research station SBI, Cuddalore and Regional Agricultural Research Station, Anakappale, AP; Agricultural Research Station (ARS), Kota, Rajasthan, and advanced sugarcane breeding clones of PAU, Ludhiana, along with commercial checks. The experiment was conducted at Sugarcane research farm Ladhawal, PAU, Ludhiana, from 2004-05 to 2006-07 as per the guidelines of AICRP on sugarcane entomology. Each genotype was raised in a plot measuring 13.5 m<sup>2</sup> with 3 replications in Randomized Block Design. All the recommended package of practices of PAU, Ludhiana, for cultivation of sugarcane crop except for any insecticidal applications were followed. Parameters recorded to assess population of stalk borer at harvest were % incidence and % intensity on the basis of 25 canes/replication. Observations were recorded based on the holes in the stalk and the internal damaged portion and infestation index was calculated on the basis of following formulae

$$\text{Infestation index} = \frac{\% \text{ incidence} \times \% \text{ intensity}}{100}$$

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## Results and Discussion

Stalk borer incidence varied widely among different genotypes tested in the present study as shown in Table 1. On the basis of overall comparison, no location-specific conclusions can be drawn which may be due to diverse parentage of test genotypes within the location. Moreover, the number of genotypes at one location is too less to draw conclusive interpretation (Table 2). But individual genotype information on stalk borer incidence is very useful to assess their potential for further utilization.

The comparison of the whole set of genotypes indicated wide variation for stalk borer incidence i.e. from 1.0% in Co Se 00421 and Sel 18-01 to 32.91% in Co S 02258. To accommodate the high level of variation, the genotypes were grouped into six different classes at an interval of 5% and designated as 1.0-5.0, 5.01-10.0, 10.01-15.0, 15.01-20.0, 20.01-25.0 and more than 25.01%. Thirty-three genotypes were present in the lowest incidence group of 1 to 5%, while four genotypes, namely, Co 98017, Co S 02258, Co S 88230 and Co S 96258 were grouped in maximum incidence group of >25% incidence. Rest of the 96 genotypes were grouped in four groups viz., 42 genotypes in 5.1-10%, 40 genotypes in 10.1-15.0%, 10 genotypes in 15.1-20.0% and 4 genotypes in 20.1-25.0%. The observations indicated that most of the genotypes showed lesser incidence of stalk borer during testing, only a few genotypes showed higher level of incidence in these three years 2004 to 2007 (Table 3).

Similarly, the infestation index also varied from 0 in Co Se 00421 and Sel 18-01 to 6.45 in CoS 02258. Forty genotypes showed infestation index < 0.5, 30 genotypes were in the range of 0.5-1.0, 40 genotypes were in the range of 1.0-2.0, 13 genotypes were in the range of 2.0-3.0 and 10 genotypes showed infestation index > 3.0.

The variability in the incidence of stalk borer in different genotypes has also been reported in the earlier studies. In a field study conducted in Punjab, stalk borer incidence ranged from zero to 6.7% and 17 genotypes (CoH92, CoLk91238, CoLk91239, CoJ87, CoH92201, CoH92202, Co88042, S465-94, CoS93227, S8846-93, S50-94, S93-94, S992-94, S161-94, CoP96219, S126-93 and CoS8436) were rated comparatively resistant to stalk borer (Anonymous, 1999). Similarly, among the 24 sugarcane varieties tested for stalk borer incidence at Nayagarh, CoC91061 found to be moderately

resistant whereas, the rest of 23 varieties were susceptible to this pest (Jena *et al.*, 1996).

In the present study genotypes CoS 8436 and CoS 767 behaved as moderately resistant against stalk borer. Similar findings have also been reported by Jaipal (1992) who found Co S 767 to be resistant to this pest amongst various sugarcane accessions and commercial hybrids evaluated.

Similarly in another screening of 15 sugarcane genotypes against *C. auricilius*, Co1148 had the highest level of stalk borer damage (7.4%) while S85-303 had the lowest (1.3%) and the genotypes S85-303 and S85-293 were least susceptible to stalk borer damage (Singh *et al.*, 1995).

The available pest management tools are inadequate to contain the losses caused by the pest. Hence, in sugarcane ecosystem varietal resistance could be a practical tool for pest management that can be fitted in the integrated pest management systems of the crop. Besides this, the resistant varieties having low pest population help in reducing the high cost of plant protection.

The promising 33 genotypes or elite clones identified in the present study are of a great significance in varietal development in sugarcane to confer insect pest resistance. Studies on parents and progeny derived from parents exhibiting a range of resistance levels for sugarcane borer (*Diatraea saccharalis*) revealed that mean percent borer-damaged internodes was higher among progeny derived from susceptible x susceptible crosses (22.8% in 1998 and 9.5%) compared to those derived from resistant x resistant crosses (20.8% in 1998 and 5.1%). Crossing a susceptible parent to a resistant parent also reduced mean percent borer-damaged internodes among the progeny compared to the susceptible parent or the susceptible x susceptible progeny and produced progeny showing transgressive segregation in both directions (Kimbeng *et al.*, 2004). Thus, the resistant genotypes identified in the present study can be further used in breeding programmes to genetically enhance resistance in sugarcane cultivars against this serious pest which can go a long way adding profitability and sustainability to the sugarcane cultivation.

A more concerted effort involving several cycles of screening and recurrent selection for resistance to borer damage is needed to identify germplasm that can withstand high levels of borer infestation.

**Table 1. Reaction of different genotypes/elite clones/varieties against stalk borer at Ludhiana**

S.No	Genotype	Mean incidence (%)	Mean intensity (%)	Infestation index
1	BO 128	6.46	5.20	0.34
2	BO 130	6.40	4.90	0.31
3	BO 91	4.41	5.12	0.23
4	Co 00120	4.61	12.92	0.60
5	Co 00237	4.27	7.63	0.33
6	Co 00238	6.19	11.11	0.69
7	Co 00239	6.99	9.34	0.65
8	Co 00240	8.65	19.55	1.69
9	Co 00241	6.44	14.68	0.95
10	Co 0116	7.32	7.76	0.57
11	Co 0117	11.84	9.88	1.17
12	Co 0118	11.10	10.55	1.17
13	Co 0121	11.70	10.02	1.17
14	Co 0122	7.14	7.79	0.56
15	Co 0123	15.87	15.06	2.39
16	Co 0124	9.73	7.81	0.76
17	Co 1148	11.73	16.09	1.89
18	Co 86249	19.34	13.80	2.67
19	Co 89003	24.10	22.16	5.34
20	Co 89029	4.51	4.66	0.21
21	<b>Co 97009</b>	13.08	15.64	2.05
22	Co 98014	4.00	7.10	0.28
23	Co 98017	28.36	12.99	3.68
24	Co 99015	23.75	15.66	3.72
25	Co 99016	13.01	17.21	2.24
26	Co A 93081	11.68	21.39	2.50
27	Co C 90063	9.75	21.35	2.08
28	Co H 110	12.26	14.22	1.74
29	Co H 115	8.38	5.43	0.46
30	Co H 117	8.79	12.72	1.12
31	Co H 118	13.38	13.46	1.80
32	Co H 119	18.81	20.07	3.78
33	<b>Co H 92201</b>	21.15	8.10	1.71
34	Co H 92202	4.23	7.69	0.33
35	Co J 64	13.12	11.66	1.53
36	<b>Co J 83</b>	9.53	8.53	0.81
37	Co J 85	6.72	11.26	0.76
38	Co J 88	11.69	11.54	1.35
39	Co J 89	2.38	1.80	0.04
40	<b>Co J 99192</b>	16.58	13.53	2.24
41	Co Lk 9412	9.84	8.93	0.88
42	Co Lk 9606	2.42	3.11	0.08
43	Co Lk 9616	10.18	17.05	1.74
44	Co Lk 9705	5.16	5.16	0.27
45	Co Lk 9707	3.61	2.55	0.09
46	Co Lk 9709	4.30	5.01	0.22
47	Co Lk 9710	6.76	8.21	0.55
48	Co Lk 97169	10.81	9.11	0.98
49	Co Pant 01215	6.80	12.99	0.88
50	Co Pant 01216	9.57	10.50	1.00
51	Co Pant 02217	7.61	12.86	0.98
52	Co Pant 02218	7.81	10.11	0.79
53	Co Pant 84211	10.88	5.11	0.56
54	Co Pant 84212	8.14	7.99	0.65
55	Co Pant 90223	7.24	6.01	0.44
56	Co Pant 99213	12.65	8.89	1.12
57	Co Pant 99214	10.79	8.86	0.96
58	Co Pk 1122	9.21	8.22	0.76
59	Co Pk 174	6.06	15.49	0.94
60	Co Pk 59	7.56	2.80	0.21
61	Co Pk 78	8.41	7.59	0.64
62	Co S 00221	8.21	15.26	1.25
63	Co S 00257	12.05	11.12	1.34
64	Co S 01256	12.10	8.81	1.07
65	Co S 01268	9.89	19.35	1.91

Contd.

Table 1 Contd.

S.No	Genotype	Mean incidence (%)	Mean intensity (%)	Infestation index
66	Co S 02252	5.40	6.50	0.35
67	Co S 02258	32.91	19.59	6.45
68	Co S 02264	9.23	12.19	1.13
69	<b>Co S 767</b>	9.35	6.22	0.58
70	Co S 8436	12.29	4.97	0.61
71	Co S 88230	26.90	18.90	5.08
72	Co S 94257	11.02	12.10	1.33
73	Co S 95258	2.62	1.11	0.03
74	Co S 96258	28.33	12.86	3.64
75	Co S 96275	12.93	14.61	1.89
76	Co S 97248	7.15	6.86	0.49
77	Co S 98259	4.66	8.44	0.39
78	Co Se 00421	1.00	0.10	0.001
79	Co Se 92423	1.85	0.66	0.01
80	Co Se 95422	10.32	9.63	0.99
81	Sel 761-98	11.74	12.34	1.45
82	Sel 842-98	10.55	15.50	1.64
83	Sel 906-98	2.13	5.74	0.12
84	Sel 922-98	10.45	14.74	1.54
85	Sel 928-98	14.21	12.61	1.79
86	Sel 943-98	11.93	10.62	1.27
87	Sel 95-98	9.77	12.70	1.24
88	Sel 17-00	10.89	16.61	1.81
89	Sel 18-00	8.21	6.10	0.50
90	Sel 19-00	2.90	0.35	0.01
91	Sel 21-00	6.60	0.89	0.06
92	Sel 22-00	6.92	5.60	0.39
93	Sel 24-00	4.41	0.69	0.03
94	Sel 25-00	11.18	18.82	2.10
95	Sel 28-00	19.20	13.87	2.66
96	Sel 49-00	16.44	14.72	2.42
97	Sel 56-00	19.49	11.84	2.31
98	Sel 63-00	2.63	0.90	0.02
99	Sel 64-00	9.36	10.17	0.95
100	Sel 67-00	6.95	16.92	1.18
101	<b>Sel 68-00</b>	7.55	16.43	1.24
102	Sel 69-00	4.65	9.11	0.42
103	Sel 70-00	3.35	10.94	0.37
104	Sel 77-00	8.11	8.50	0.69
105	Sel 11-01	10.73	12.44	1.33
106	Sel 12-01	17.06	25.28	4.31
107	Sel 17-01	11.65	8.85	1.03
108	Sel 18-01	1.00	0.10	0.001
109	Sel 56-01	17.48	14.65	2.56
110	Sel 61-01	2.32	25.00	0.58
111	Sel 69-01	10.16	15.12	1.54
112	Sel 1046-03	10.01	8.23	0.82
113	Sel 1059-03	10.50	11.44	1.20
114	Sel 112-03	20.10	19.56	3.93
115	Sel 113-03	9.32	23.45	2.19
116	Sel 140-03	1.14	2.10	0.02
117	Sel 145-03	1.44	0.50	0.01
118	Sel 147-03	1.33	2.36	0.03
119	Sel 149-03	10.51	9.58	1.01
120	Sel 152-03	11.05	12.21	1.35
121	Sel 164-03	1.04	0.31	0.003
122	Sel 174-03	20.00	22.80	4.56
123	Sel 187-03	10.01	10.23	1.02
124	Sel 281-03	1.23	2.90	0.04
125	Sel 33-03	10.10	10.80	1.09
126	Sel 49-03	2.03	3.20	0.06
127	Sel 50-03	1.50	2.50	0.04
128	Sel 589-03	3.10	6.30	0.20
129	Sel 806-03	2.40	5.68	0.14
130	Sel 902-03	10.10	10.89	1.10
131	Sel 91-03	4.26	22.41	0.95
132	Sel 945-03	10.40	18.90	1.97

**Table 2. Regional distribution of genotypes evaluated against stalk borer at Ludhiana**

S.No.	Location	Series	Total No. of genotypes evaluated	<5 % incidence		>15 % incidence	
				No.	Per cent	No.	Per cent
1	Bihar and Orissa	BO	3	1	33.3	0	0
2	Tamil Nadu (Coimbatore)	Co	22	4	18.18	5	22.73
3	Andhra Pradesh (Anakapalli)	Co A	1	0	0	0	0
4	Kerala (Cuddalore)	Co C	1	0	0	0	0
5	Haryana (Karnal)	Co H	7	1	14.29	2	28.57
6	Punjab (Jalandhar)	Co J	6	1	16.67	1	16.67
7	UP (Lucknow)	Co Lk	8	3	37.5	0	0
8	Uttarakhand (PantNagar)	Co Pant	9	0	0	0	0
9	Rajasthan (Kota)	Co Pk	4	0	0	0	0
10	UP (Shahjahanpur)	Co S	16	2	12.5	3	18.75
11	UP (Seorahi)	Co Se	3	2	66.67	0	0
12	Punjab (Ludhiana 1998)	Sel	7	1	14.29	0	0
13	Punjab (Ludhiana 2000)	Sel	17	5	29.41	3	17.65
14	Punjab (Ludhiana 2001)	Sel	7	2	28.57	2	28.57
15	Punjab (Ludhiana 2003)	Sel	21	10	47.62	2	9.52
Total			132	32	24.24	18	13.63

**Table 3. Percent incidence of Stalk borer in different genotypes**

Range (%)	No. of genotypes
1.0-5.0	33
5.01-10.0	42
10.01-15.0	40
15.01-20.0	10
20.01-25.0	4
>25.01	4

## References

- Anonymous (1999) Annual report of sugarcane entomology for the year 1998-99. Punjab Agricultural University, Ludhiana, India.
- Avasthy PN (1983) Insect Pest Management for Sugarcane in India. In: M Balasubramaniam and AR Solayappan (eds), *Sugarcane Pest Management in India*. Tamil Nadu Cooperative Federation, Madras, pp 71-77.
- Chaudhary JP, SC Bhardwaj and JN Gupta (1980) Failure of insecticidal sprays for the control of stalk borer, *Chilo auricilius* Ddgn. *Indian J. Agric. Res.* **14**: 22-28.
- Duhra MS and KS Suri (2002) Insects and their control. In: Sidhu GS and KS Waraitch, *Sugarcane Production and Protection Technologies*. Punjab Agricultural University, Ludhiana, India, pp 85-106.
- Jaipal S (1992) Susceptibility of some sugarcane accessions and cultivars to major moth borers. *Tests Agrochemicals and Cultivars* **13**: 120-121.
- Jaipal S (2000) An IPM module for the management of major insect pests of sugarcane in Indian subtropics. *Sugar Tech.* **2(3)**: 1-8.
- Jayanthi R and YS Goud (2002) Sugarcane pests: borers. In: SB Singh, G Pyrilla Rao and S Easwaramoorthy (eds) *Sugarcane Crop Management* SCI Tech Publishing LLC, Teas USA, pp 530-551.
- Jena BC, NC Patnaik and AK Parida (1996) Intensity of stalk borer attack in sugarcane. *Indian Sugar* **46**: 537-540.
- Kimbeng CA, WH White, JD Miller and BL Legendre (2004) Sugarcane resistance to the sugarcane borer: performance among progeny derived from resistant and susceptible parents. *J. Am. Soc. Sugarcane Technol.* **24**: 81-104.
- Singh D, SR Yadav, H Singh and DS Ahlawat (1995) Screening of sugarcane genotypes against stalk borer *Chilo auricilius* Ddgn. infestation under natural conditions. *Haryana J. Agron.* (<http://www.cababstractsplus.org/abstracts.AcNo=19951603582>).